

Molecular Characterization and Genetic Diversity of *Entamoeba* spp. from Free-ranging Macaque in Thailand

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► Introduction

- *Entamoeba* is protozoan
 - Phylum Sarcomastigophora
 - Subphylum Sarcodina



Figure 1 *Entamoeba histolytica/dispar*. SAF-fixed specimen, wet mount examination. Above, an elongated trophozoite with two nuclei (telophase). Below, a rounded one (pre-cystic form)

- Cause of **dysentery and invasive disease** in animals and human (Hoare, 1959).

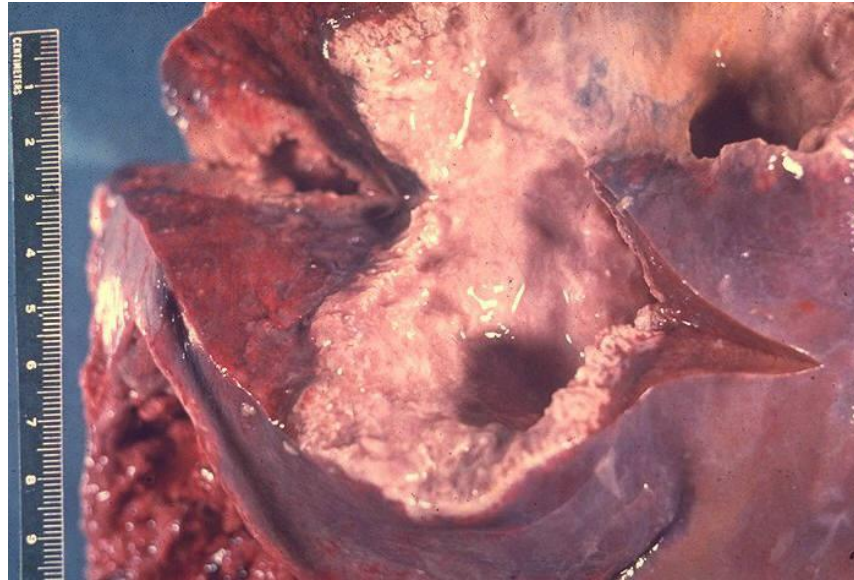


Figure 3 Gross pathology of liver containing amebic abscess

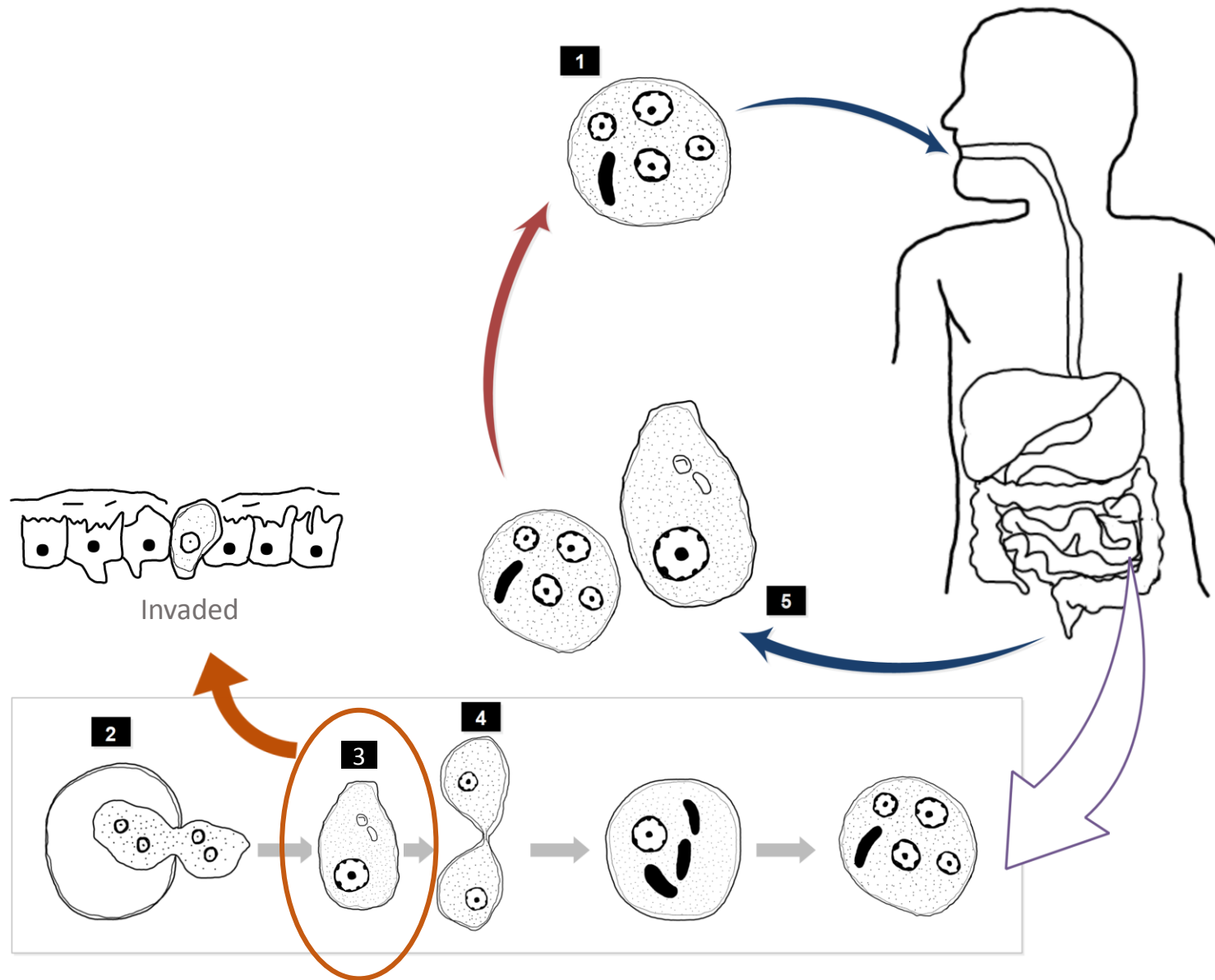


Figure 2 Life cycle of *Entamoeba* (1) *Entamoeba histolytica* infections occur by ingestion of mature cysts in contaminated food, water, or dirty hands. (2) Cyst go through small intestine and excyst to be trophozoite. (3) Trophozoites migrate to large intestine. (4) Trophozoites multiply by asexual replication (binary fission) and encyst. (5) Trophozoites and cysts were excreted with feces.

- The six species of *Entamoeba* can be found in **human** feces

- *Entamoeba histolytica*

- *Entamoeba dispar*
- *Entamoeba moshkovskii*
- *Entamoeba hartmanni*
- *Entamoeba coli*
- *Entamoeba polecki*

← Pathogenic →

- *Entamoeba histolytica*

- *Entamoeba nuttali*
- *Entamoeba dispar*
- *Entamoeba hartmanni*
- *Entamoeba coli*
- *Entamoeba polecki*
- *Entamoeba chattoni*

- *E. nuttali*; In Belgium and Netherlands had the report of *E. nuttali* infection in zookeeper who work with the infectious animals (Levecke et al., 2015).

► Epidemiology

- This disease spread worldwide, commonly found in the developing countries (Acha & Szyfres, 2001; Samad, 2013).
- From 2007 to 2011, the samples of returned traveler were collected and showed positive results with *E. histolytica* from the travelers, most of them came back from Mexico, India, Indonesia, or Thailand (“Amebiasis - Chapter 3 - 2016 Yellow Book | Travelers’ Health | CDC,” n.d.).

- In 2015 from the annual epidemiology surveillance report showed 10.78 of dysentery cases per 100,000 inhabitants, may has cause from amoebic 42.58%.
- Most of cases were found in the border area and north part of Thailand, especially in remote rural area.
- The highest incidence occurred in May and June period, and lowest incidence occur in September. (Bureau of Epidemiology, Department of Disease Control, n.d.).

► Transmission

- The **ingestion of cyst** contaminated in food or water, especially the raw vegetables and undercooked food (Acha & Szyfres, 2001; Samad, 2013).
- **Oral-anal sexual** (Lim & Vythilingam, 2013; Tanyuksel & Petri, 2003; Ximénez et al., 2009).
- **Globalization** (Kruse & others, 2004; Schuster & Visvesvara, 2004).

► Zoonosis

- Especially in non-human primates
 - They have similar genetic with human
 - they are known as naturally harbor of *Entamoeba* with the same strains in human infection.

It demonstrated by the experiment in 1934 about the cross-infection of *E. histolytica* in simian strains and human strains (*Advances in Parasitology*, 1967; Hoare, 1959).

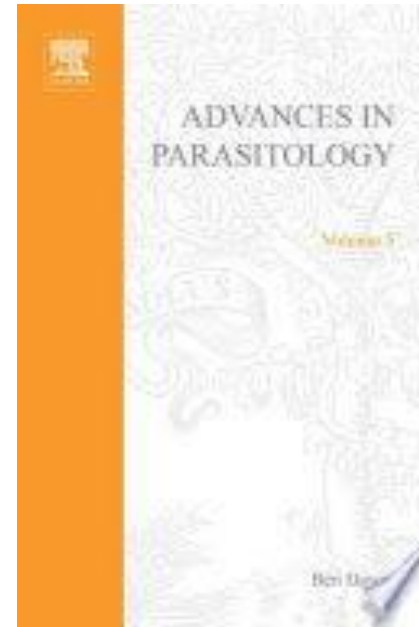


Figure 4 *Advances in Parasitology*, 1967

“The transmission of zoonotic diseases often occurs when human and animals coexist and share the resources together”
(Lane-deGraaf et al., 2014)



Figure 5 Increasing of macaque population in Lopburi



Figure 6 Monkey buffet in Thailand

► Diagnosis

- Molecular method
 - Gold standard (Fletcher et al., 2012; Tanyuksel & Petri, 2003).
 - High sensitivity and specificity more than other method.
 - It can detect pathogens from a variety of clinical specimens.
 - Need the specialized equipment, technically staff, and high costs
- The 18s rRNA is widely used as gene target for Entamoeba identification because of the ubiquitous and conserved within species (Fotedar et al., 2007).

► Project

► Objectives of study

- To examine pathogenic *Entamoeba* spp. exist in monkey population in Lopburi province, Thailand.
- To establish the study on molecular characterization and genetic diversity of *Entamoeba* spp. infection in NHPs.

► Material and Methods

- The **456** free-ranging macaque stools were collected from the **city center** and **Erawan non-hunting area**, Lopburi province, Thailand.
- The samples were preserved in D-solution and kept at 4°C
- Extraction by the phenol-chloroform method and storing at -20°C until use.



Figure 7 *Macaca leonina* in the community around Erawan non-hunting area, Lopburi province



Figure 8 Sample collection points at City center area, Lopburi province, Thailand

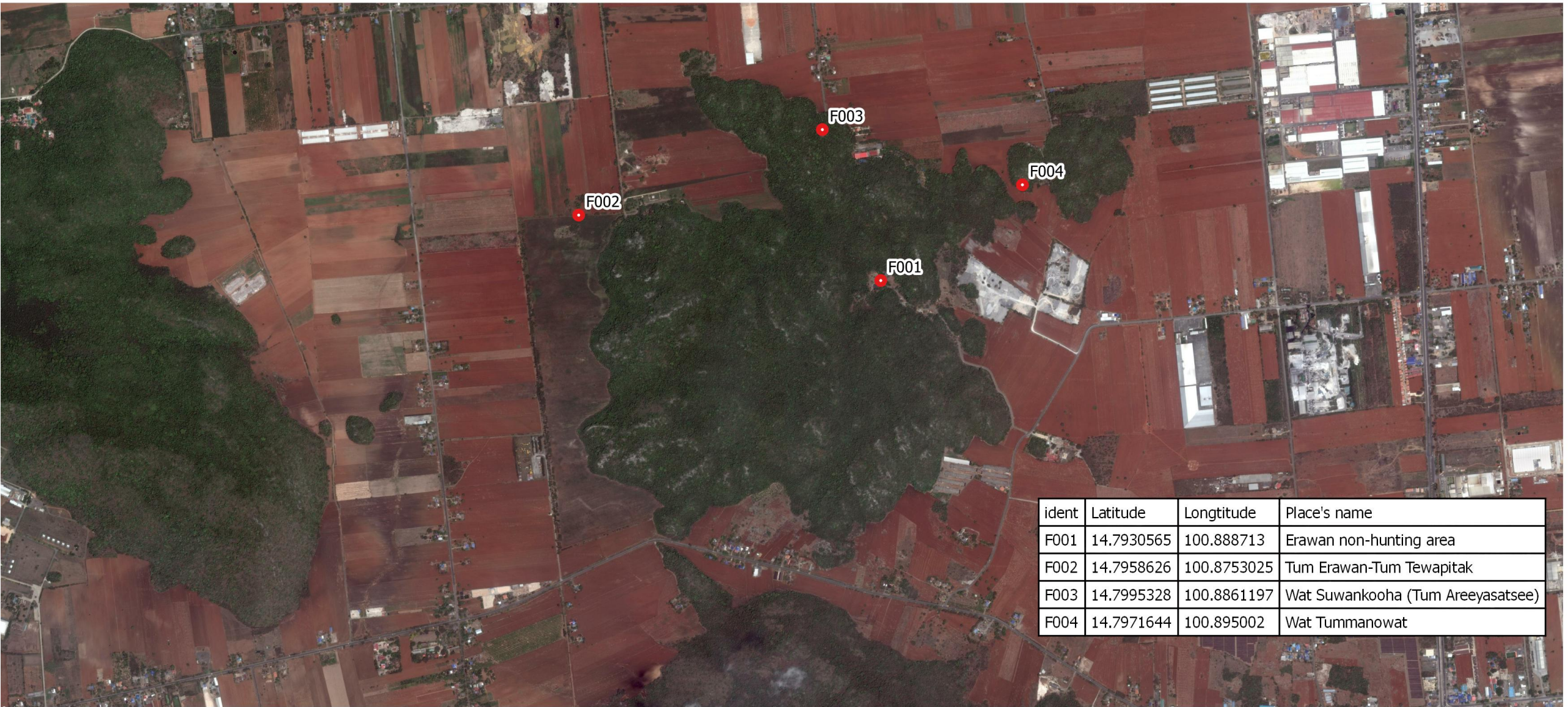


Figure 9 Sample collection points at Erawan non-hunting area, Lopburi province, Thailand

- PCR by using forward primer Entam1 and reverse primer Entam2 (Table 1) with pre-denature 3 minutes at 95°C, follow by 40 cycles of 40 seconds at 94°C, 30 seconds at 50°C, 30 seconds at 72°C for, and the last extension 5 minutes at 72°C.

Table 1 Primers use for the conventional PCR for *Entamoeba* spp. (Suzuki, Kobayashi, Murata, Yanagawa, & Takeuchim, 2007)

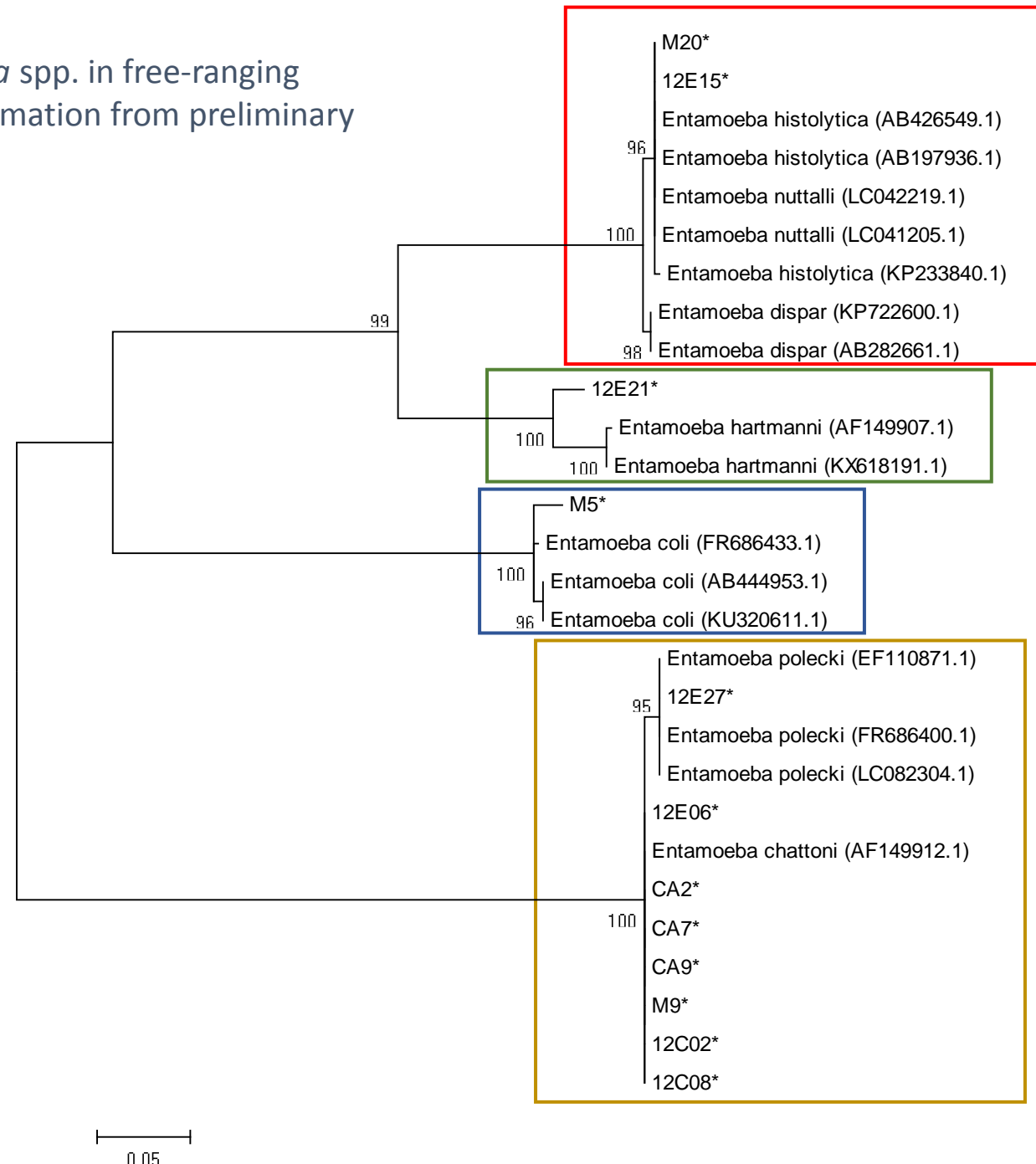
Primers	Sequences	Amplification product (bp)
Entam1(Fw)	5'- GTT GAT CCT GCC AGT ATT ATA TG-3'	506
Entam2(Rw)	5'-CAC TAT TGG AGC TGG AAT TAC-3'	

- The positive samples were done again with the new forward primer (18s-entam-ku-1w) and three new reverse primers (18s-entam-poc/chat-ku-2w, Entam-coli-2w, and 18s-entam-path-ku-2w) (Table 2).
- PCR parameters were started by 3 minutes at 98°C, follow by 40 cycles of 40 seconds at 98°C, 30 seconds at 58°C, 30 seconds at 72°C for, and the last extension 5 minutes at 72°C.

Table 2 The new forward and reverse primers for specific group of *Entamoeba* spp.

Primers	Sequences	Amplification product (bp)	
18s-entam-ku-1w	5'-ATC TGG TTG ATC CTG CCA GTA TT-3'		
18s-entam-poc/chat-ku-2w	5'-CCT TCA AGT TGA TAG GTC AGA TAT TTA AAA CAG-3'	315	
Entam-coli-2w	5'-CAC TAC CTC CTT CAG TTG TTA AGA T-3'	516	
18s-entam-path-ku-2w	5'-CAT TTT GTA CTA ATA CAA ACT GGA TCG TC-3'	199	

Figure 10 Phylogenetic tree of *Entamoeba* spp. in free-ranging macaques based on 18S rRNA gene (information from preliminary study)



► Result and Discussion

- The 190 from 456 fecal samples showed positive results with primer Entam1 and Entam2.
- All of those were detected again with new specific primers. The result showed 170 positive samples that separated to 8 group results (Table 3).

Table 3 The result of new specific primers

	Number of samples
<i>E. polecki</i> or <i>E. chattoni</i> (group I)	40
<i>E. coli</i> (group II)	4
<i>E. histolytica</i>, <i>E. nuttali</i>, or <i>E. dispar</i> (group III)	1
Coinfection with group I and II	30
Coinfection with group I and III	37
Coinfection with group II and III	4
Coinfection with group I, II, and III	54
Negative with new primers	18

New primers

- The negative results of new primers showed the sequencing of plant, fungi, and *E. hartmanni*
 - The new primers are specific more than the old primer

Future plan

- The specific primer for *E. hartmanni* group should be created and add in the set of new specific primers.
- Need more sequencing results and phylogenetic tree
- Analyze the information
- Publish



THANK YOU
FOR YOUR ATTENTION